## A Coupled Oscillator Calculation of the Induced Circular Dichroism of the Visual Pigment Analog (S)-N-all-trans-Retinylidene-α-(1-naphthyl)ethylamine<sup>†</sup>

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ABSTRACT: The visual pigment rhodopsin exhibits a Cotton effect in the region of 500 nm. The preceding paper (Johnston, E. M., and Zand, R. (1973), *Biochemistry 12*, 4631) describes the synthesis and CD absorption spectra of (S)-N-all-transretinylidene- $\alpha$ -(1-naphthyl)ethylamine, which resembles rhodopsin in possessing a Cotton effect in its main long-wavelength absorption band. Whether a twisted chromophore mechanism (intrinsic mechanism) or a coupled oscillator mechanism (extrinsic mechanism) can best account for the observed optical activity is an unresolved question. In an attempt to answer the question, the rotational strengths to be

expected from a coupled oscillator mechanism were calculated for (S)-N-all-trans-retinylidene- $\alpha$ -(1-naphthyl)ethylamine using the Kirkwood theory. All of the calculated rotational strengths agree within a factor of two with those found experimentally. Therefore, the 490-nm Cotton effect of rhodopsin is not *prima facie* evidence for the distortion of the retinaldehyde chromophore. Instead the retinaldehyde transitions could become optically active by coupling with the transitions of aromatic amino acid side chains located within a 15-Å radius of the chromophore.

In the preceding paper (Johnston and Zand, 1973) we reported the preparation and the absorption and circular dichroism (CD) spectra of a number of Schiff's base visual pigment analogs derived from 11-cis- or all-trans-retinaldehydes and simple optically active amines. Having demonstrated that such simple amine moieties are capable of inducing optical activity into the conjugated chromophoric system of the retinaldehyde molecule, it became desirable to see if a mechanism for the process could be established. The mechanism proposed by Crescitelli and coworkers (1966) for the extrinsic Cotton effect in rhodopsin requires the presence of an imposed geometric asymmetry on the retinaldehyde moiety resulting in an intrinsic mechanism for the origin of the observed CD. Mommaerts (1969) has argued that 11-cis-retinaldehyde can assume one of two possible enantiomeric forms and that the observed circular dichroism arises through the preferential stabilization of one of these forms by the protein opsin. Some support for the Mommaerts position has recently appeared and is derived from theoretical considerations (Nash, 1969; Honig and Karplus, 1971; Honig et al., 1973), nuclear magnetic resonance (Honig et al., 1971), and the Xray crystal structure determination of 11-cis-retinaldehyde (Gilardi et al., 1971).

Rhodopsins containing isomers other than the 11-cis isomer also exhibit optical activity. Isorhodopsin (Takezaki and Kito, 1967) contains the 9-cis-retinaldehyde isomer while metarhodopsin contains the all-trans isomer and is also optically active (Waggoner and Stryer, 1971). Honig et al. (1973) have suggested that the planar chain isomers are twisted about the 6-7 bond and if one enantiomer of the twisted ring-chain

bond is selected for the iso- and metarhodopsins then the observed optical activity can be explained by an intrinsic mechanism. In the case of our visual pigment analogs it is difficult to conceive of a mechanism that would preferentially select and stabilize one particular conformer of the *all-trans*-retinaldehyde.

A coupled oscillator mechanism (Kirkwood, 1937) would seem to be a suitable first choice to explain both the rhodopsin data and our visual pigment analog data. To carry through such a calculation we chose to utilize the spectra obtained from the model compound (S)-N-all-trans-retinylidene- $\alpha$ -(1-naphthyl)ethylamine, ((S)-RNEA). A coupled oscillator approach has been successfully used to explain the CD of myoglobin (Hsu and Woody, 1969, 1971). In the ideal situation, it would be desirable to perform a similar calculation for either rhodopsin or our model compound. This is not feasible at this time since the X-ray structures have not been determined for either of these substances whereas the X-ray structure for myoglobin is known. Ultimately one might be able to locate proximal aromatic side chains by an analysis of the CD of several transitions of a chromophore. For rhodopsin there are Cotton effects at 340 and 490 nm. If a reliable calculation method was available, these two Cotton effects might yield information about the environment of the chromophore. The rationale for the present calculation was that it might eliminate the coupled oscillator mechanism on an order of magnitude basis if the predicted CD was much weaker than the experimentally observed CD. Not only the rotational strengths but also the wavelengths and signs of the bands could be tested by this approach.

Method of Calculation. In carrying through such a calculation it is preferable to use a rigid molecule for the analysis. Otherwise, the calculated CD must be averaged over the allowed conformations, which introduces an additional component of uncertainty. Rigidity is not possible for a Schiff's

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<sup>&</sup>lt;sup>1</sup> Abbreviation used is: (S)-RNEA, (S)-N-all-trans-retinylidene- $\alpha$ -(1-naphthyl)ethylamine.

FIGURE 1: The rotation angles  $\phi_1$ , the N-C $_{\alpha}$  rotation, and  $\phi_2$ , the C $_{\alpha}$ -C $_1$  rotation, in a molecule of (S)-N-all-trans-retinylidene- $\alpha$ -(1-naphthyl) ethylamine.

base of retinaldehyde. However, steric hindrances may restrict the allowed conformation into a narrow range. In our calculation we sought to produce two maps. The first map would show the conformational energy as a function of the two rotation angles shown in Figure 1. The second map would show the rotational strength of a particular induced band as a function of these two angles. Thus, the CD spectrum is predictable from the sterically allowed conformations. If some flexibility remains, in spite of the steric restrictions, one may try to average the rotational strength over the allowed regions.

Hard Sphere Map. As a preliminary test of our method, a hard sphere map for (S)-RNEA was calculated. The bond lengths and bond angles used are given in Table II. The nonbonded contact distances used are given in Table II. The two rotation angles  $\phi_1$ , the N-C<sub> $\alpha$ </sub> rotation, and  $\phi_2$ , the C<sub> $\alpha$ </sub>-C<sub>1</sub> rotation, are indicated in Figure 1 and each of these angles is defined so that  $\phi = 0$  in the (S)-cis conformation. A positive

TABLE I: Bond Lengths and Angles Used in the Calculation.

Retinylidene Group						
C == C	1.36 Å)					
C-C	1.46 Å	Inuzuka and Becker (1968)				
All C-C-C-	120°					
angles						
Naphthyl Group						
$C_i = C_2$	1.365 Å					
$C_1 = C_9$	1.425 Å	Abrahams et al. (1949), as re-				
$C_9 = C_{10}$	1.393 Å	calculated by Ahmed and				
$C_2 = C_3$	1.404 Å	Cruickshank (1952)				
<C <sub>9</sub> C <sub>10</sub> C <sub>5</sub>	120°	Assumed				

These five quantities completely determine the structure if one assumes  $D_{2h}$  symmetry. The other two angles will differ slightly from  $120^{\circ}$ .

Ethylamine Linkage  $N-C_{\alpha}$ 1.48 Å Sutton (1958), p S-16 (from  $CH_3N=NCH_3$ )  $C_{\alpha}$ --- $C_{\text{methyl}}$ 1.54 Å Sutton (1958)  $C_{\alpha}-C_{\text{ring}}$ 1.52 Å Sutton (1958), p S-13 (from toluene) C-H 1.08 Å Sutton (1958) 120°  $< C_{14} - C_{15} = N$ Bürgi and Dunitz (1969) give 122.7° for N-benzylideneaniline and 120.2° for N-pmethylbenzylidene-p-nitroaniline 120°  $< C_{15} = N - C_{\alpha}$ Bürgi and Dunitz (1969) give 119.9° for N-benyzlideneaniline 109.47° All angles at Assumed tetrahedral  $C_{\alpha}$  and  $C_{\text{methyl}}$ 

TABLE II: Nonbonded Contact Distances Used for the Hard-Sphere Map of (S)-RNEA (Figure 3).<sup>a</sup>

	We will be a second of the sec
$C \cdots H$	2.20 Å
$\mathbf{N}\cdots\mathbf{H}$	2.20 Å
$H \cdot \cdot \cdot H$	1.90 Å
$\mathbf{C} \cdot \cdot \cdot \cdot \mathbf{C}$	3.00 Å

<sup>&</sup>lt;sup>a</sup> These distances are the inner limits given by Ramakirshnan and Ramachandran (1965).

rotation turns the group at the far end clockwise as seen from the near end. The convention employed in that of Sugeta and Miyazawa (1967). A computer program was written in Fortran IV to calculate the position of each relevant atom as a function of the two rotation angles. Successive coordinate transformations using the transformation matrices of Sugeta and Miyazawa (1967) were used to carry out this calculation.

The retinylidene group is assigned its own coordinate system as is the naphthyl group. The positions of the atoms within each group are fixed by the bond lengths and angles of Table I. The resulting coordinates for the retinylidene and naphthyl atoms are given in Tables III and IV. A syn configuration at the C to N double bond was assumed.

Conformational Energy Map. An improvement on the hardsphere calculation is to use a conformational energy calculation. The nonbonded potential energy functions are those of Bartell (1966) except for one change. The H···H function was modified slightly for computer use, to avoid a spurious minimum at r = 0. The functions used are mostly repulsive and do not include the electrostatic attractions that have been used in many polypeptide calculations such as those of Brant and Flory (1965). Most of the conformations that are forbidden by the energy map have very high energies and the inclusion of electrostatic attractions would not alter the outer limits significantly. Also, there is some doubt whether the attractive forces in such a highly conjugated system can be accurately represented as a sum of pairwise interactions. The attractive forces between polyenes calculated by Haugh and Hirschfelder (1955) cannot be reduced to a sum of pairwise

Calculation of Rotational Strengths. For a retinylidene transition at frequency  $\nu_R$  and a given naphthyl transition of frequency  $\nu_N$ , the rotational strength is given by the equation

TABLE III: Coordinates of Selected Retinylidene Atoms."

Atom	X	<i>T</i> ,	Atom	X	<i>h</i> .
C <sub>5</sub>	0.00000	0.00000	C <sub>12</sub>	8.49173	0.70397
$C_6$	1.16363	-0.70397	$C_{13}$	9.77080	0.00000
$\mathbb{C}_7$	2.44270	0.00000	$C_{14}$	10.93443	-0.70397
$C_8$	3.60633	-0.70397	$C_{15}$	12.21350	0.00000
C <sub>9</sub>	4.88540	0.00000			
$C_{10}$	6.04903	-0.70397	$H_{14}$	10.93129	-1.78397
C <sub>11</sub>	7.32810	0.00000	$\mathbf{H}_{15}$	12.21664	1.08000

<sup>a</sup> See Figure 2 for atom numbers. The origin of the retinylidene coordinate system is at  $C_5$ . The positive x axis passes through  $C_7$ ,  $C_9$ ,  $C_{41}$ ,  $C_{13}$ , and  $C_{15}$ . The xy plane is chosen to pass through the remaining retinylidene atoms, with the positive y axis pointing away from  $C_6$ . Coordinates are given in angstroms.

TABLE IV: Coordinates of the Naphthyl Atoms.

Atom	x	у	Atom	x	у
$C_{i}$	0.0000	0.0000	$H_2$	0.1670	-2.1029
$C_2$	0.7070	-1.1676	$H_3$	2.6510	-2.1029
$C_3$	2.1110	-1.1676	$H_4$	3.8980	0.0000
$C_4$	2.8180	0,0000	$H_5$	3.8980	2.4682
$C_5$	2.8180	2.4682	$H_6$	2.6510	4.5711
$C_6$	2.1110	3.6358	$H_7$	0.1670	4.5711
$\mathbb{C}_7$	0.7070	3.6358	$H_8$	-1.0800	2.4682
$C_8$	0.0000	2.4682			
C <sub>9</sub>	0.7125	1.2341			
$C_{10}$	2.1055	1.2341			

<sup>a</sup> See Figure 2 for atom numbers. The origin of the naphthyl coordinate system is at  $C_1$ . The xy plane is the plane of the naphthyl group. The positive x axis passes through  $C_4$ ; the positive y axis passes through  $C_3$ . Coordinates are in angstroms.

$$R = \frac{2\pi}{C} \frac{V_{\text{OR;ON}} \nu_{\text{R}} \nu_{\text{N}} \mathbf{R}_{\text{RN}} \cdot (\mathbf{y}_{\text{R}} \times \mathbf{y}_{\text{N}})}{h(\nu_{\text{N}}^2 - \nu_{\text{R}}^2)}$$
(1)

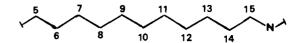
where  $V_{\rm OR;ON}$  is the coulomb matrix element between the retinylidene transition and the naphthyl transition (Tinoco, 1962). That is

$$V_{\mathrm{OR;ON}} = \iint \phi_{\mathrm{RO}}(\mathbf{r}_1) \phi_{\mathrm{R_1}} \frac{2e^2}{\tau_{12}} \phi_{\mathrm{N_0}}(\mathbf{r}_2) \phi_{\mathrm{N_1}}(\mathbf{r}_2) d\tau_1 d\tau_2 \qquad (2)$$

For our purposes all the group wave functions can be taken to be real. Equation 1 is valid for the interaction of two nondegenerate, electrically allowed, magnetically forbidden transitions, where the interaction energy V is a small perturbation on the total energy of the groups. The transition dipole moments and frequencies were obtained from the experimental spectra. The dipole moment of all-trans-retinaldehyde was calculated to be 9.67 D by integrating the 360-nm peak of the absorption spectrum of all-trans-retinaldehyde in isooctane. The dipole moments for the three naphthyl transitions were calculated from the oscillator strengths reported by Klevens and Platt (1949). The evaluation of  $V_{\rm OR;ON}$  was carried out by using the monopole approximation of London (1942) and free-electron molecular orbital (FEMO) wave functions for the ground and excited state for both the retinylidene group and the naphthyl group.

FEMO Wave Functions. The original shape of the polyene was used for these calculations rather than the flattened out polyene representation of Haugh and Hirschfelder (1955). The conjugated chain of the retinylidene group includes carbons  $C_5$  through  $C_{15}$ , plus the nitrogen atom so the chain of twelve atoms is an even atom chain. The single bond lengths are taken as 1.46 Å and the carbon–carbon double bond lengths are taken as 1.36 Å. According to Salem (1966), even atom chains should be extended by half a bond length at each end to obtain the length of the bond. If 1.27 Å is allowed for the length of the C=N—bond and if the box is extended by 1.46 Å/2 at one end and 1.27 Å/2 at the other end the total length of the box is 16.84 Å. The shape of the one-dimensional box is shown in Figure 2. The FEMO orbitals for this box are

$$\phi_n(x) = \sqrt{\frac{2}{16.84}} \sin\left(\frac{n\pi x}{16.84}\right) \text{ for } n = 1, 2, \dots 6 \quad (3)$$



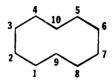


FIGURE 2: The one-dimensional box representation of the retinylidene and naphthyl moieties of (S)-N-all-trans-retinylidend- $\alpha$ -(1-naphthyl)ethylamine.

where x is a length coordinate that follows the box rather than going in a straight line.

The ground state of the retinylidene can be constructed as a Slater determinant over the above six orbitals, containing the coordinates of twelve electrons. Then, the first excited state involving the promotion of one electron from the 6th to 7th orbital is an  $f \rightarrow g$  transition, in Platt's notation (Platt, 1961), and the electric dipole moment for this transition reduces to

$$\mathbf{u}_{01} = \sqrt{2}e \int \phi_6 \mathbf{r} \phi_7 d\tau \tag{4}$$

which involves single-particle orbitals rather than Slater determinants; formulas for this reduction are given by Murrell (1963).

Transition Densities. It is convenient to define a transition density for the  $\psi_0 \rightarrow \psi_1$  transition of the retinylidene, namely,  $\rho_{01} = (2)^{1/2} e \phi_6 * \phi_7$ . Since  $\phi_6$  is real here, this becomes  $\rho_{01} = (2)^{1/2} e \phi_6 \phi_7$ . Substitution into eq 4 results in the expression

$$\mathbf{u}_{\mathrm{Ro}_{1}} = \int \rho_{\mathrm{R}_{0}} \mathbf{r} \mathrm{d}\tau \tag{5}$$

For a given naphthyl transition a similar expression can be written

$$\mathbf{u}_{N_{01}} = \int \rho_{N_{01}} \mathbf{r} d\tau \tag{6}$$

These transition densities may be substituted into eq 2 for the coulomb matrix element to yield:

$$V_{\text{OR;ON}} = \int \int \rho_{\text{Ro}}(\mathbf{r}_1) \frac{2e^2}{r_{12}} \rho_{\text{No}}(\mathbf{r}_2) d\tau_1 d\tau_2$$
 (7)

The value of the coulomb matrix element is thus reduced to the interaction energy between two "transition charge" distributions.

Monopole Approximation. The monopole approximation of London is an approximation of the matrix element V as a sum of the interactions of point charges, or transition monopoles:

$$V_{\text{OR;ON}} = \sum_{j} \sum_{k} \frac{G_{\text{R}j} G_{\text{N}k}}{r_{\text{R}j\text{N}k}}$$
 (8)

where  $G_{Rj}$  is the jth monopole of the retinylidene group,  $G_{Nk}$  is the kth monopole of the naphthyl group, and  $r_{RjNk}$  is the distance between the two monopoles.

Evaluation of Retinylidene Monopoles. Since the retinylidene

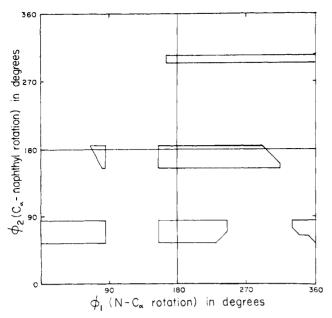


FIGURE 3: The hard sphere map calculated with  $C_{15}$ —N= $C_{\alpha}$  equal to  $114^{\circ}$  and the  $C_{\alpha}$  methyl group represented as a bare carbon atom.

group has been represented as a zigzag box along the polyene skeleton with a kink at every carbon atom, the transition charge density  $\rho_{\rm R}(x)=(2)^{1/2}e\phi_6(x)\phi_7(x)$ , contains the length coordinate x that follows the box rather than going in a straight line. The "nodal surfaces" used in the monopole approximation are here points, the zeros of the function

$$\rho = \frac{2\sqrt{2}}{l}e\sin\frac{6\pi x}{l}\sin\frac{7\pi x}{l} \tag{9}$$

TABLE V: FEMO Monopoles for Retinvlidene Group.

	Charge		
No.	(Electrons)	x (Å)	y (Å)
1	0.027176	-0.16868	-0.09284
2	0.176380	0.53488	-0.32359
3	0.006785	1.24954	-0.65669
4	-0.002575	1.61564	-0.45519
5	0.043836	2.23807	-0.11263
6	0.127419	2.85238	-0.24785
7	-0.000781	3.57773	-0.68667
8	-0.018380	3.88191	-0.55230
9	0.048898	4.65655	-0.12595
10	0.063873	5.15141	-0.16093
11	-0.022651	2.89131	-0.60855
12	-0.034450	6.27365	-0.58034
13	0.040112	7.07000	-0.14205
14	0.016988	7.45866	-0.07898
15	-0.072713	8.19374	-0.52369
16	-0.040058	8.69331	-0.59302
17	0.019131	9.43704	-0.18369
18	0.000029	9.77526	-0.00270
19	-0.136853	10.48686	-0.43320
20	-0.034403	11.11565	-0.60423
21	0.002575	11.69140	-0.28735
22	-0.011298	12.10649	-0.05890
23	-0.179217	12.81203	-0.36210
24	-0.019826	13.43931	-0.56051

The dipole moment of this transition charge distribution is parallel to the long axis of the retinylidene group.

In order to locate the monopoles, the integration of every peak between a pair of successive zeros was modified by the addition of cuts in the charge distribution at the position of the atom, *i.e.*, kinks. The integral  $\int_{C_i}^{C_i^{i+1}} \rho(x) dx$  was taken between every pair of successive cuts,  $C_i$  and  $C_i^{i+1}$ , and a monopole was placed at the center of gravity of each segment. By this procedure all of the monopoles were placed on the polyene skeleton. The position and charges of the retinylidene monopoles are given in Table V. Before being used in the rotational strength calculation, the monopole strengths were multiplied by a factor so that the dipole moment of the whole monopole distribution became equal to the experimental dipole moment for the transition being considered.

Evaluation of Naphthyl Monopoles. For naphthalene, the one-dimensional FEMO box is taken as the perimeter. The  $C_9$ – $C_{10}$  bridge is neglected. The box is kinked as with the retinylidene group, but now runs in a closed loop around the carbon skeleton as illustrated in Figure 3.

The free electron orbitals to be used for the naphthyl group have the same form as those used for benzene by Salem (1966). The ten naphthyl electrons will go into the lowest five orbitals, those for which  $\Delta=0,\pm1,$  and  $\pm2.$ 

The singlet excited state wave functions can be obtained as combinations of Slater determinants in the following way

$$\psi_{1} = \frac{1}{\sqrt{2}} (\chi_{2 \to 3} + \chi_{-2 \to -3})$$

$$\psi_{2} = \frac{1}{\sqrt{2}} (\chi_{2 \to 3} - \chi_{-2 \to -3})$$

$$\psi_{3} = \frac{1}{\sqrt{2}} (\chi_{2 \to -3} + \chi_{-2 \to 3})$$

$$\psi_{4} = \frac{1}{\sqrt{2}} (\chi_{2 \to -3} + \chi_{-2 \to 3})$$

where

$$\chi_{2 o 3} = rac{1}{\sqrt{2}} \{ \left| \phi_{-2} ar{\phi}_{-2} \phi_{-1} ar{\phi}_{-1} \phi_{0} ar{\phi}_{0} \phi_{1} ar{\phi}_{1} \phi_{2} ar{\phi}_{3} \right| + \left| \phi_{-2} ar{\phi}_{-2} \cdot \cdot \cdot \cdot \phi_{3} ar{\phi}_{2} \right| \}$$

and  $\psi_0$  is the ground state.

The transition densities for these electrons can be obtained using the following expressions

For

$$\psi_0 = \frac{1}{\sqrt{2}}(\chi_{j\to k} + \chi_{-j\to -k})$$

$$\rho = \sqrt{2}e\left(\frac{1}{\sqrt{2}}\right)(\phi_j^*\phi_k + \phi_{-j}^*\phi_{-k})$$

where

$$\phi_j = \frac{1}{\sqrt{2\pi}} e^{ij\theta}$$

$$\rho = \frac{e}{2\pi} [e^{i\theta(k-j)} + e^{-i\theta(k-j)}]$$

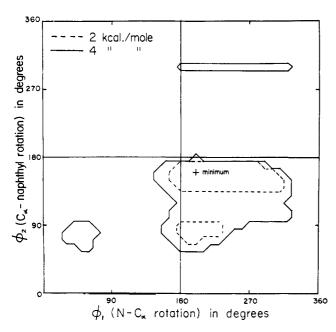


FIGURE 4: The conformational energy map calculated with  $C_{15}$ — $N=C_{\alpha}$  equal to 120° and the  $C_{\alpha}$  methyl group represented as an actual methyl group.

when j = 2, k = 3

$$\rho = \frac{e \cos \theta}{\pi}$$

If the zero point for  $\theta$  is taken at  $C_{10}$ , then this density function has loops at the atoms, and this is the  ${}^{1}B_{a}$  transition density. By similar methods we obtained the other three transition densities. The four expressions are

$$\rho_{^{1}B_{a}} = \frac{e \cos \theta}{\pi}$$
short axis polarized
$$\rho_{^{1}L_{a}} = \frac{e \sin 5 \theta}{\pi}$$

$$\rho_{^{1}B_{b}} = \frac{e \sin \theta}{\pi}$$

$$\log \text{ axis polarized}$$

$$\rho_{^{1}L_{b}} = \frac{e \cos 5 \theta}{\pi}$$

The <sup>1</sup>L<sub>b</sub> transition was not included in the calculation because of its low intensity.

The procedure for computing naphthyl charges and locations, starting from the transition densities was the same as that described for the retinylidene group. The positions and charges of these monopoles are listed in Table VI.

## Results

The hard-sphere map resulting from our calculations and embodying the restrictions listed in Table II is shown in Figure 4. The molecular geometry given in Table I was used, with the following two exceptions. The angle  $C_{15}$ —N= $C_{\alpha}$  was set equal to 114° and the methyl group on  $C_{\alpha}$  was represented as a bare carbon atom. The energy was calculated for all combi-

TABLE VI: FEMO Monopoles for the Naphthyl Group.

No.	Charge (Electrons)	<i>x</i> (Å)	y (Å)
	1. <sup>1</sup> L <sub>2</sub> T	ransition	
1	0.098363	0.53935	0.93419
2	0.091986	0.18678	0,32352
3	0.067169	0.16472	-0.27203
4	0.045053	0.50868	-0.84007
5	0.015739	0.94178	-1.16760
6	-0.015739	1.87622	-1.16760
7	-0.045053	2.30932	-0.84007
8	-0.067169	2.65328	-0.27203
9	-0.091986	2.63122	0.32352
10	-0.067169	2.27865	0.93419
11	-0.098363	2.27865	1.53401
12	-0.091986	2.63122	2.14468
13	-0.067169	2.65328	2.74023
14	-0.045053	2.30932	3.30827
15	-0.015739	1.87622	3,63580
16	0.015739	0.94178	3.63580
17	0.045053	0.50868	3.30827
18	0.067169	0.16472	2.74023
19	0.091986	0.18678	2.14468
20	0.098363	0.53935	1.53401
			1,00.01
1	0.063186	Transition 0.24080	0.41708
1 2	0.156271	0.24080	-0.62698
3	0.197707	1.40900	-1.16760
4	0.156271	2.43836	-0.62698
5	0.063186	2.43830	
6			0.41708
7	-0.063186	2.57720	2.05112
	-0.156271	2.43836	3,09518
8	-0.197707	1.40900	3.63580
9	-0.156271	0.37964	3.09518
10	-0.063186	0.24080	2.05112
		Transition	0. 63000
1	0.190349	0.36897	0.63908
2	0.112222	0.30281	-0.50008
3	0.015739	0.94178	<b>-1.16760</b>
4	-0.015739	1.87622	-1.16760
5	-0.112222	2.51519	-0.50008
6	-0.190349	2.44903	0.63908
7	-0.190349	2.44903	1.82912
8	-0.112222	2.51519	2.96828
9	-0.015739	1.87622	3.63580
10	0.015739	0.94178	3,63580
11	0.112222	0.30281	2.96828
12	0.190349	0.36897	1.82912

nations  $(\phi_1, \phi_2)$  using ten-degree increments. If the hard-sphere map is valid, the steric restrictions are rather severe and only about 10% of the map area is permitted.

The conformational energy map used for the CD calculations is shown in Figure 5. It is based on the potential functions of Table VII and uses a value of  $120^{\circ}$  for the  $C_{15}$ —N= $C_{\alpha}$  rather than  $114^{\circ}$  as was used in the hard sphere map. Also the methyl group on  $C_{\alpha}$  was represented as such rather than a bare carbon atom. It was allowed to rotate freely to its minimum energy position for each pair of values given to  $\phi_1$  and  $\phi_2$ . The region inside the 4-kcal/mol contour represents about 15% of the area of the map. There is an allowed area

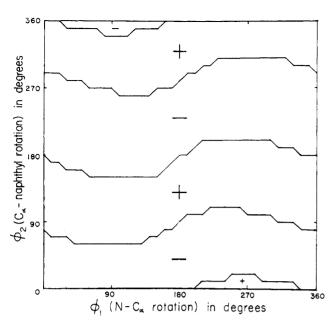


FIGURE 5: The sign of the calculated rotational strength as a function of  $\phi_1$  and  $\phi_2$  for the f  $\rightarrow$  g (360-nm) band in the retinylidene group.

in the lower right quadrant, a smaller area to its left and a very narrow slot in the upper right quadrant at  $\phi_2 = 300^{\circ}$ .

The calculated rotational strength of the 360-nm transition of the retinylidene group can be mapped as a function of the angles as shown in Figure 5. The map shows the sign of this rotational strength as a function of  $\phi_1$  and  $\phi_2$ . It is obtained by summing the contributions of the three naphthyl transitions,  ${}^1\!L_a$ ,  ${}^1\!B_b$ ,  ${}^1\!B_a$ . Sign maps for the induced rotational strengths of the three naphthyl transitions are given in Figures 6 and 7.

The 360-nm transitions of the retinylidene group was observed to have a positive rotational strength. If this fact is used to restrict the allowed pairs of angles  $(\phi_1, \phi_2)$  then about 50% of the map area is not allowed. A further restriction is that the 282-nm induced band of the naphthyl was observed to have a negative rotational strength. This restricts  $(\phi_1, \phi_2)$  to lie in the region of Figure 6 marked with a negative sign. Lastly, the CD induced in the  ${}^{1}B_b$  transition at 224 nm was observed to be positive. Superposition of Figures 5, 6, and 7 shows that the signs are given correctly only if  $(\phi_1, \phi_2)$  lies in a narrow twisting region reaching across the map from

TABLE VII: Electric Transition Dipole Moments Used in the Calculation."

Retinylidene band	9.669 D	
Naphthyl bands		
$^{1}\mathrm{L}_{\mathrm{a}}$	3.326	
$^1\mathrm{B_b}$	8.923	
$^{1}\mathrm{B}_{\mathrm{a}}$	4.619	

<sup>a</sup> These dipole moments come from observed adsorption spectra and were used to calculate the theoretical CD spectrum of (S)-RNEA. The value for the retinylidene transition was obtained by integrating the peak in the *all-trans*-retinal absorption spectrum. The values for the naphthyl transitions were obtained from the oscillator strengths for naphthalene given by Klevens and Platt (1949).

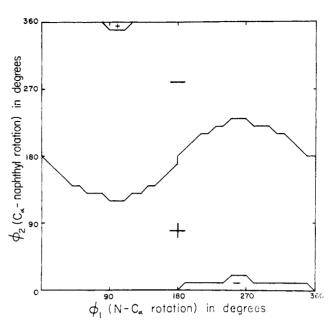


FIGURE 6: Sign of the calculated rotational strengths as a function of  $\phi_1$  and  $\phi_2$  for the naphthyl  ${}^1L_a$  (282-nm) and  ${}^1B_a$  bands.

left to right and centered at  $\phi_1 = 180^\circ$  and  $\phi_2 = 300^\circ$ . The region includes about 5% of the map area. Comparison of these results with the energy map of Figure 4 reveals that no point in the large low-energy region of the lower right quadrant would give the correct set of signs for the three rotational strengths. The only permissible region is the narrow low-energy strip in the upper right quadrant, reaching from (180°, 300°) to (320°, 300°). The points in this region are all between 2 and 4 kcal per mol above the global minimum energy. Only the points (180°, 300°), (190°, 300°), and (200°, 300°) give the correct signs for the three rotational strengths. The conformation (180°, 300°) has been selected as the one that gives the best agreement with the observed spectrum. Theoretical rotational strengths (180°, 300°) are compared graphically in Figure 8.

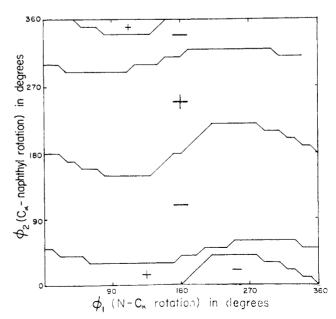


FIGURE 7: Sign of the calculated rotational strengths as a function of  $\phi_1$  and  $\phi_2$  for the naphthyl  ${}^1B_b(224\text{-nm})$  band.

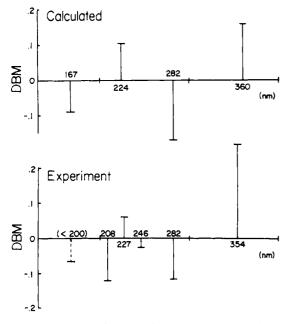


FIGURE 8: Comparison of the calculated and experimental values of the induced rotational strengths, wavelengths, and signs of (S)-N-all-trans-retinylidene- $\alpha$ -(1-naphthyl)ethylamine.

## Discussion

Our calculation shows that there exists a conformation for (S)-N-all-trans-retinylidene- $\alpha$ -(1-naphthyl)ethylamine that predicts four rotational strengths within a factor of two, and gives all their signs correctly. The  $^1B_n$  rotational strength taken from the spectrum was an estimate only, since it was obtained as the negative of the sum of the observed rotational strengths. However, it does agree in sign and approximate magnitude with the calculated  $^1B_n$  rotational strength. The wavelengths of the three observed CD bands also agree within 6 nm with the locations of the transitions they are predicted to result from.

An a priori prediction of the CD spectrum without having the observed spectrum would have led to a choice of the minimum energy conformation, which lies in the lower right quadrant of the energy map at  $(200^{\circ}, 160^{\circ})$ . This choice would have resulted in the wrong sign for both the  $^{1}L_{a}$  and  $^{1}B_{b}$  predicted rotational strengths. Restricting the conformation to the region of the energy map reading from  $(180^{\circ}, 300^{\circ})$  to  $(320^{\circ}, 300^{\circ})$  would not improve the results obtained. If a Boltzmann average were performed over this allowed region the sign of the retinylidene band would come out wrong, since for the majority of this region the sign is negative.

The fact that reasonable agreement, between the calculated and observed CD spectra, results for a particular conformation is encouraging. The specific factors that cause the conformation of (S)-RNEA to be fixed at (180°, 300°) are not known at this time. It may be possible to obtain some answers to this question by applying the method to other molecules having similar conformational restrictions.

The results of this calculation in conjunction with the experimental study in the previous paper (Johnston and Zand, 1973) show that the 490-nm Cotton effect of rhodopsin cannot

be viewed as arising necessarily from an imposed distortion of the polyene chain of retinaldehyde. A major contributing mechanism if not the sole mechanism, for this Cotton effect could be the interaction of the polyene transitions with the transitions of nearby (less than 15 Å) aromatic side chains of the protein opsin.

## References

Abrahams, S. C., Robertson, J. M., and White, J. G. (1949), Acta Crystallogr. 2, 238.

Ahmed, F. R., and Cruickshank, D. W. J. (1952), Acta Crystallogr. 5, 852.

Bartell, L. S. (1960), J. Chem. Phys. 32, 827.

Brant, D. A., and Flory, P. J. (1965), J. Amer. Chem Soc. 87, 2791.

Bürgi, H. B., and Dunitz, J. D. (1969), Chem. Commun., 472.

Crescitelli, F., Mommaerts, W. F. H. M., and Shaw, T. I. (1966), *Proc. Nat. Acad. Sci. U. S.* 56, 1729.

Gilardi, R., Karle, I., Karle, J. and Sperling, W. (1971), Nature (London) 232, 187.

Haugh, E. F., and Hirschfelder, J. O. (1955), *J. Chem. Phys.* 23, 1778.

Honig, B., Hudson, B., Sykes, B., and Karplus, M. (1971), *Proc. Nat. Acad. Sci. U. S.* 68, 1289.

Honig, B., Kahn, P., and Ebrey, T. G. (1973), *Biochemistry* 12, 1637.

Honig, B., and Karplus, M. (1971), Nature (London) 229, 556.Hsu, M. C., and Woody, R. W. (1969), J. Amer. Chem. Soc. 91, 3679.

Hsu, M. C., and Woody, R. W. (1971), J. Amer. Chem. Soc. 93, 3515

Inuzuka, K., and Becker, R. S. (1968), *Nature (London) 219*, 383

Johnston, E. M., and Zand, R. (1972), Biochem. Biophys. Res. Commun. 47, 713.

Johnston, E. M., and Zand, R. (1973), *Biochemistry* 12, 4631. Kirkwood, J. G. (1937), *J. Chem. Phys.* 5, 479.

Klevens, H. B., and Platt, J. R. (1949), *J. Chem. Phys. 17*, 470. London, F. (1942), *J. Phys. Chem. 46*, 305.

Mommaerts, W. F. H. M. (1969), in The Retina: Morphology, Function and Clinical Characteristics, Straatsma, B. R., Hall, M. D., Allen, R. A., and Crescitelli, F., Ed., Los Angeles, Calif., University of California Press, p 225.

Murrell, J. N. (1963), The Theory of the Electronic Spectra of Organic Molecules, New York, N. Y., Wiley.

Nash, H. A. (1969), J. Theoret. Biol. 22, 314.

Platt, J. R. (1961), in Handbuch der Physik, Flugg, S., Ed., Vol. 37, Berlin, Springer, p 173.

Ramakrishnan, C., and Ramachandran, G. N. (1965), Biophys. J. 5, 909.

Salem, L. (1966), The Molecular Orbital Theory of Conjugated Systems, New York, N. Y., Benjamin.

Sugeta, H., and Mijazawa, T. (1967), Biopolymers 5, 673.

Sutton, L. E., Ed. (1958), Interatomic Distances and Configurations in Molecules and Ions, London, Chemical Society.

Takezaki, M., and Kito, Y. (1967), *Nature (London) 215*, 1197. Tinoco, I., Jr. (1962), *Advan. Chem. Phys. 4*, 113.

Waggoner, A. S., and Stryer, L. (1971), Biochemistry 10, 3250.